

Preliminary communication

Stereocontrolled synthesis of sialyl Lewis X ceramide consisting of a pentasaccharide recognized by the selectin family *

Akira Hasegawa, Takashi Ando, Akihiko Kameyama and Makoto Kiso

Department of Applied Bioorganic Chemistry, Gifu University, Gifu 501-11 (Japan)

(Received February 11th, 1992; accepted March 12th, 1992)

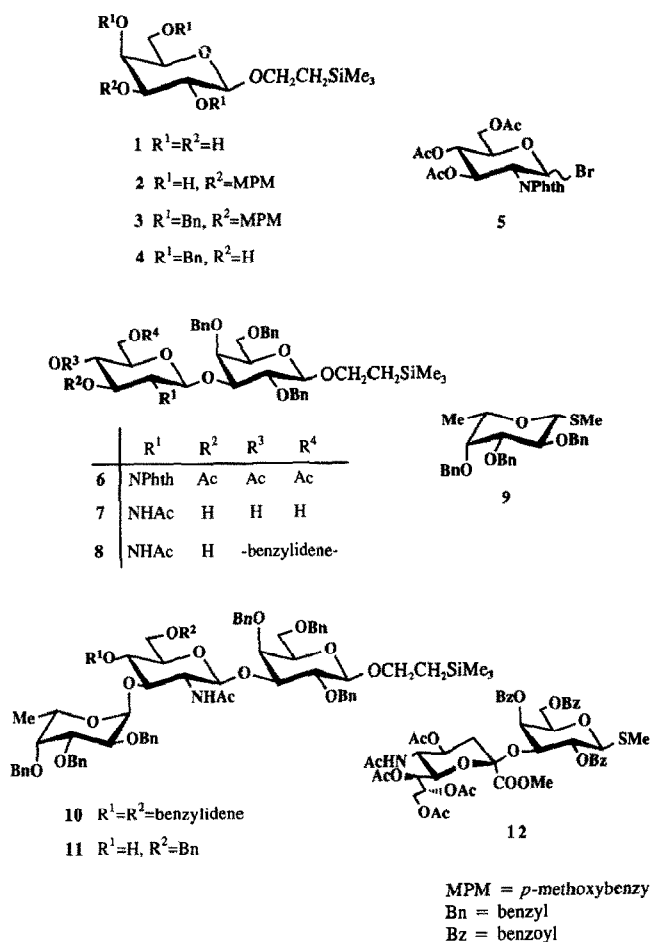
Glycoconjugates containing sialic acid have received much attention owing to their important functions in biological systems². Very recently, it has been demonstrated³ that the selectin family, such as E-selectin (endothelial leukocyte adhesion molecule-1, ELAM-1), P-selectin (GMP-140), and L-selectin (LECAM-1), recognizes the sialyl Le^X determinant, α -Neu5Ac-(2 $\rightarrow$ 3)- β -D-Gal-(1 $\rightarrow$ 4)-[α -L-Fuc-(1 $\rightarrow$ 3)]- β -D-GlcNAc, which is found as the terminal carbohydrate structure in both glycolipids and glycoproteins.

Previously, we have reported the synthesis of sialyl Le^X ganglioside⁴ (a hexasaccharide), sialyl (2 $\rightarrow$ 6) α Le^X ganglioside⁵, and analogues¹, and have examined recognition activity by E-selectin. The data indicated that both the fucose and sialic acid residues were required for recognition and sialyl (2 $\rightarrow$ 6) α Le^X ganglioside was not recognized. In view of these facts, it is of interest to elucidate the more detailed structural requirements necessary for selectin recognition. We describe herein the stereocontrolled synthesis of sialyl Le^X ceramide, α -Neu5Ac-(2 $\rightarrow$ 3)- β -D-Gal-(1 $\rightarrow$ 4)-[α -L-Fuc-(1 $\rightarrow$ 3)]- β -D-GlcNAc-(1 $\rightarrow$ 3)- β -D-Gal-(1 $\rightarrow$ 1)-ceramide, which is a pentasaccharide.

The glycosylation (Scheme 1) of 2-(trimethylsilyl)ethyl 2,4,6-tri-*O*-benzyl- β -D-galactopyranoside (4), prepared from 2-(trimethylsilyl)ethyl β -D-galactopyranoside (1, ref. 6) via dibutyltin oxide-mediated, selective 3-*O*-4-methoxybenzylation, *O*-benzylation, and oxidative removal of the 4-methoxybenzyl group with 2,3-dichloro-5,6-dicyanobenzoquinone⁷, was effected with 2,4,6-tri-*O*-acetyl-2-deoxy-2-phthalimido-D-glucopyranosyl bromide⁸ (5). Reaction in the presence of silver carbonate, silver perchlorate, and powdered 4A molecular sieves (MS-4A) in

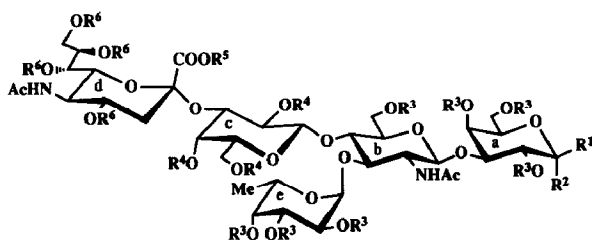
Correspondence to: Professor A. Hasegawa, Department of Applied Bioorganic Chemistry, Gifu University, Gifu 501-11, Japan.

* Synthetic Studies on Sialoglycoconjugates, Part 39. For Part 38, see ref. 1.



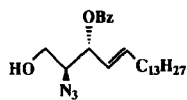
Scheme 1.

dichloromethane gave a 76% yield of the desired β -glycoside **6** {mp 140–142°, $[\alpha]_D -4.5^\circ$ (CHCl_3)}, showing in its $^1\text{H-NMR}$ spectrum a one-proton doublet at δ 5.81 ($J_{1,2}$ 8.4 Hz, H-1), characteristic of the 2-phthalimido- β -D-glucopyranosyl unit. *O*-Deacetylation of **6**, followed by heating with hydrazine hydrate in 95% aqueous ethanol, and *N*-acetylation of the product afforded 2-(trimethylsilyl)ethyl *O*-(2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 $\rightarrow$ 3)-2,4,6-tri-*O*-benzyl- β -D-galactopyranoside (**7**) {mp 94–96°, $[\alpha]_D -22^\circ$ (CHCl_3)} in 82% yield. Treatment of **7** with benzaldehyde dimethyl acetal in the presence of *p*-toluenesulfonic acid gave the benzylidene derivative **8** in 88% yield. Compound **8** was used as the glycosyl acceptor. The glycosylation of **8** with methyl 2,3,4-tri-*O*-benzyl-1-thio- β -L-fucopyranoside⁹ (**9**) in benzene for 10 h at 5–10°, in the presence of dimethyl(methylthio)sulfonium triflate (DMTST)¹⁰ and MS-4A, afforded the desired trisaccharide **10** $[\alpha]_D -63^\circ$ (CHCl_3) in 97% yield, showing in its $^1\text{H-NMR}$ spectrum a three-proton doublet at δ 0.87 ($J_{5,6}$ 6.4 Hz, CH_3CH) and a one-proton

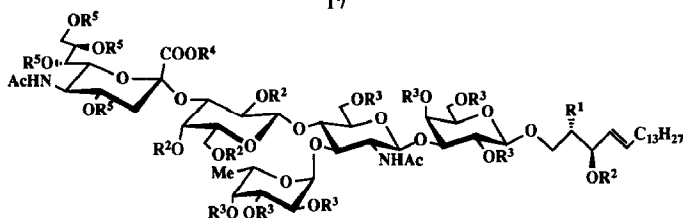


	R ¹	R ²	R ³	R ⁴	R ⁵	R ⁶
13	OSE	H	Bn	Bz	Me	Ac
14	OSE	H	Ac	Bz	Me	Ac
15		OH, H	Ac	Bz	Me	Ac
16	H	OC(=NH)CCl ₃	Ac	Bz	Me	Ac

SE = 2-(trimethylsilyl)ethyl



17



	R ¹	R ²	R ³	R ⁴	R ⁵
18	N ₃	Bz	Ac	Me	Ac
19	NHCOC ₁₇ H ₃₅	Bz	Ac	Me	Ac
20	NHCOC ₁₇ H ₃₅	H	H	H	H

Scheme 2.

doublet at δ 5.07 ($J_{1,2}$ 3.6 Hz, H-1), characteristic of the α -L-fucopyranosyl unit. Reductive ring-opening of the benzylidene group in **10** with sodium cyanoborohydride–hydrogen chloride in dry tetrahydrofuran, according to the method of Garegg et al.¹¹ afforded compound **11** in 77% yield. The glycosylation of **11** with methyl *O*-(methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 $\rightarrow$ 3)2,4,6-tri-*O*-benzoyl-1-thio- β -D-galactopyranoside¹² (**12**), in dichloromethane for 16 h at 0° in the presence of DMTST and MS-4A, gave a 57% yield of the pentasaccharide **13** $\{[\alpha]_D -26^\circ$ (CHCl₃) $\}$, which had the desired stereochemistry (Scheme 2). Significant signals in the ¹H-NMR spectrum of **13** were a three-proton doublet at δ 1.15 ($J_{5,6}$ 6.4 Hz, CH₃CH, fucose unit), two three-proton singlets, at δ 1.58 and 1.61 (*N*-COCH₃), four three-proton singlets, at δ 1.80, 1.93, 1.98, and 2.15 (OCOCH₃), a three-proton singlet at δ 3.79

(OCH₃), and a complex resonance at δ 7.14–8.22 (7 C₆H₅), a one-proton doublet of doublets at δ 5.46 ($J_{1,2}$ 8.1, $J_{2,3}$ 10.1 Hz, H-2 of the benzoylated Gal unit), indicating the newly formed glycosidic linkage to be β -D. Removal of the benzyl groups from **13** by catalytic hydrogenolysis over 10% Pd/C in 5:1 ethanol–formic acid for 48 h at 45°, and subsequent acetylation gave compound **14** $\{[\alpha]_D -24^\circ$ (CHCl₃) $\}$ in 66% yield. Selective removal of the 2-(trimethylsilyl)ethyl group from **14** by treatment¹³ with trifluoroacetic acid in dichloromethane for 30 min at 0° gave compound **15** in quantitative yield. Treatment¹⁴ of **15** with trichloroacetonitrile in the presence of 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) for 4 h at 0° gave the corresponding α -trichloroacetimidate **16** $\{[\alpha]_D +6^\circ$ (CHCl₃) $\}$ in 85% yield. Significant signals in the ¹H-NMR spectrum were a one-proton doublet at δ 6.49 ($J_{1,2}$ 3.8 Hz, H-1 of the acetylated Gal unit) and a one-proton singlet at δ 8.65 (C=NH), indicating α -trichloroacetimidate formation.

The final glycosylation of (2*S*,3*R*,4*E*)-2-azido-3-*O*-benzoyl-4-octadecene-1,3-diol¹⁵ **17** with **16** in dichloromethane for 8 h at 0° in the presence of boron trifluoride etherate^{14a,15a} and MS-4A, afforded only the expected β -glycoside **18** $\{[\alpha]_D -18^\circ$ (CHCl₃) $\}$, in 71% yield after column chromatography. Selective reduction¹⁶ of the azide group in **18** with hydrogen sulfide in aqueous pyridine gave the amine, and this latter compound on condensation with octadecanoic acid in the presence of 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (WSC) in dichloromethane for 24 h at room temperature afforded the sialyl Le^x ganglioside (pentasaccharide) derivative **19** $\{[\alpha]_D -9.7^\circ$ (CHCl₃) $\}$ in 80% yield. Finally, *O*-deacylation of **19** with sodium methoxide in methanol and saponification of the methyl ester group, furnished the end product **20** $\{[\alpha]_D -19^\circ$ (5:4:0.7 CHCl₃–MeOH–H₂O) $\}$ in quantitative yield after chromatography on a column of Sephadex LH-20.

In conclusion, by using compounds **5**, **9**, **12**, and **16** as glycosyl donors, and compounds **4**, **8**, **11**, and **17** as glycosyl acceptors, a stereocontrolled synthesis of sialyl Le^x ganglioside (pentasaccharide) was achieved. This ganglioside was recognized³ⁱ by E-selectin to almost the same degree as the sialyl Le^x ganglioside (hexasaccharide).

New compounds obtained gave elemental analyses and IR and ¹H-NMR (270 MHz) data in agreement with the structures assigned.

ACKNOWLEDGMENT

This work was supported in part by a Grant-in-Aid (No. 03255207) for Scientific Research on Priority Areas from the Ministry of Education, Science and Culture of Japan.

REFERENCES

- 1 H. Furui, M. Kiso, and A. Hasegawa, *Carbohydr. Res.*, submitted.
- 2 (a) S. Hakomori, *Biochem. Biophys. Acta*, 417 (1975) 55–89; (b) H. Wiegandt, in H. Wiegandt (Ed.), *New Comprehensive Biochemistry, Glycolipids*, Vol. 10, Elsevier, Amsterdam, 1985, pp. 199–260; (c)

- S. Tsuji, T. Yamakawa, M. Tanaka, and Y. Nagai, *J. Neurochem.*, 50 (1988) 414–423; (d) E.C. Bremor, J. Schlessinger, and S. Hakomori, *J. Biol. Chem.*, 261 (1986) 2434–2440.
- 3 (a) G. Walz, A. Aruffo, W. Kolanus, M. Bevilacqua, and B. Seed, *Science*, 250 (1990) 1132–1135; (b) M.J. Polley, M.L. Philips, E. Wayner, E. Nudelman, A.K. Singhal, S. Hakomori, and J.C. Paulson, *Proc. Natl. Acad. Sci. U.S.A.*, 88 (1991) 6224–6228; (c) M.L. Philips, E. Nudelman, F.C.A. Gaeta, M. Perez, A.K. Singhal, S. Hakomori, and J.C. Paulson, *Science*, 250 (1990) 1130–1132; (d) M. Tiemeyer, S.J. Swiedler, M. Ishihara, M. Moleland, H. Schweingruber, P. Hirtzer, and B.K. Brandley, *Proc. Natl. Acad. Sci. U.S.A.*, 88 (1991) 1138–1142; (e) E. Larsen, T. Palabrica, S. Sajer, G.E. Gilbert, D.D. Wagner, B.C. Furier, and B. Furie, *Cell*, 63 (1990) 467–474; (f) Y. Imai, M.S. Singer, C. Fennie, L.A. Lasky, and S.D. Rosen, *J. Cell Biol.*, 113 (1991) 1213–1221; (g) B.K. Brandley, S.J. Swiedler, and P.W. Robbins, *Cell*, 63 (1990) 861–863; (h) C. Foxall, S.R. Watson, D. Dowbenko, C. Fennie, L.A. Lasky, M. Kiso, A. Hasegawa, D. Asa, and B.K. Brandley, *J. Cell Biol.*, submitted; (i) D. Tyrrell, P. James, N. Rao, C. Foxall, S. Abbas, F. Dasgupta, M. Nashed, A. Hasegawa, M. Kiso, D. Asa, J. Kidd, and B.K. Brandley, *Proc. Natl. Acad. Sci. U.S.A.*, 88 (1991) 10372–10376; (j) M. Larkin, T.J. Ahern, M.S. Stoll, M. Shaffer, D. Sako, J. O'Brien, C.-T. Yuen, A.M. Lawson, R.A. Childs, A. Hasegawa, M. Kiso, G.R. Larsen, and T. Feizi, *J. Biol. Chem.*, submitted; (k) A. Takada, K. Ohmori, N. Takahashi, K. Tsuyuoka, A. Yago, K. Zenita, A. Hasegawa, and R. Kannagi, *Biochem. Biophys. Res. Commun.*, 179 (1991) 713–719.
- 4 (a) A. Kameyama, H. Ishida, M. Kiso, and A. Hasegawa, *Carbohydr. Res.*, 209 (1991) c1–c4; (b) A. Kameyama, H. Ishida, M. Kiso, and A. Hasegawa, *J. Carbohydr. Chem.*, 10 (1991) 546–560.
- 5 A. Kameyama, H. Ishida, M. Kiso, and A. Hasegawa, *J. Carbohydr. Chem.*, 10 (1991) 729–738.
- 6 K. Jansson, T. Frejd, J. Kihlberg, and G. Magnusson, *Tetrahedron Lett.*, 27 (1986) 753–756.
- 7 Y. Oikawa, T. Tanaka, K. Horita, T. Yoshida, and O. Yonemitsu, *Tetrahedron Lett.*, 25 (1984) 5383–5386.
- 8 R.U. Lemieux, T. Takeda, and B. Chung, *ACS Symp. Ser.*, 39 (1976) 90–115.
- 9 F. Yamazaki, T. Kitajima, T. Numata, Y. Ito, and T. Ogawa, *Carbohydr. Res.*, 201 (1990) 15–30.
- 10 (a) P. Fügedi and P.J. Garegg, *Carbohydr. Res.*, 149 (1986) c9–c12; (b) M. Ravenscroft, R.M.G. Roberts, and J.G. Tillett, *J. Chem. Soc., Perkin Trans. 2*, (1982) 1569–1572.
- 11 P.J. Garegg, H. Hultberg, and S. Wallin, *Carbohydr. Res.*, 108 (1982) 97–101.
- 12 (a) A. Kameyama, H. Ishida, M. Kiso, and A. Hasegawa, *Carbohydr. Res.*, 193 (1989) c1–c5; (b) A. Kameyama, H. Ishida, M. Kiso, and A. Hasegawa, *ibid.*, 200 (1990) 269–285.
- 13 K. Jansson, S. Ahlfors, T. Frejd, J. Kihlberg, G. Magnusson, J. Dahmen, G. Noori, and K. Stenvall, *J. Org. Chem.*, 53 (1988) 5629–5641.
- 14 (a) R.R. Schmidt and G. Grundler, *Synthesis*, (1981) 885–887; (b) M. Numata, M. Sugimoto, K. Koike, and T. Ogawa, *Carbohydr. Res.*, 163 (1987) 209–225; (c) T. Murase, A. Kameyama, K.P.R. Kartha, H. Ishida, M. Kiso, and A. Hasegawa, *J. Carbohydr. Chem.*, 8 (1989) 265–283.
- 15 (a) Y. Ito, M. Kiso, and A. Hasegawa, *J. Carbohydr. Chem.*, 8 (1989) 285–294; (b) R.R. Schmidt and P. Zimmermann, *Angew. Chem., Int. Ed. Engl.*, 25 (1986) 725–726.
- 16 (a) T. Adachi, Y. Yamada, I. Inoue, and M. Saneyoshi, *Synthesis*, (1977) 45–46; (b) T. Murase, H. Ishida, M. Kiso, and A. Hasegawa, *Carbohydr. Res.*, 188 (1989) 71–80.